### Principle of Lactophenol Cotton Blue (LPCB) Staining

Lactophenol Cotton Blue (LPCB) Staining method works on the principle of aiding the identification of the fungal cell walls.

- Fungi are eukaryotic organisms with both macroscopic and microscopic characteristics.
- The fungal spore cell wall is made up of chitin of which the components of the Lactophenol Cotton Blue solution stains for identification.
- The lactophenol cotton blue solution acts as a mounting solution as well as a staining agent.
- The solution is clear and blue in color and it is made up of a combination of three main reagents:
  - Phenol: It acts as a disinfectant by killing any living organisms
  - Lactic acid: To preserve the fungal structures
  - **Cotton blue**: To stain or give color to the chitin on the fungal cell wall and other fungal structures
- The stain will give the fungi a blue-colored appearance of the fungal spores and structures, such as hyphae.

# Reagents of Lactophenol Cotton Blue (LPCB) Staining

A preparation of 50ml Lactophenol cotton Blue staining solution is made up of:

- Distilled water 50ml
- Cotton Blue (Aniline Blue) 0.125g
- Phenol Crystals (C<sub>6</sub>H<sub>5</sub>O<sub>4</sub>) 50g
- Glycerol 100ml
- Lactic acid (CH₃CHOH COOH) 50ml
- 70% ethanol

Note: Lactophenol Cotton Blue solution is prepared at least 2 days before use.

# **Preparation of Lactophenol Cotton Blue solution**

Lactophenol Cotton Blue solution is prepared for over two days leaving the reagents undisturbed to allow dissolving and maturation.

- 1. Day 1: Dissolve the cotton blue in distilled water and leave to rest overnight. This eliminates insoluble dye.
- 2. Day2: Using protective gloves, add phenol crystals to lactic acid in a glass beaker and stir using a magnetic stirrer until the crystals dissolve.
- 3. Add glycerol

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- 4. Filter the Cotton blue and the distilled water into the phenol + glycerol +lactic acid solution and mix.
- 5. Store at room temperature.

Procedure of Lactophenol Cotton Blue (LPCB) Staining

- 1. On a clean microscopic glass slide, add a drop of 70% ethanol
- 2. Add the fungal specimen to the drop of alcohol using a sterile mounter such as an inoculation loop (from solid medium), depending on the sample of use.
- 3. Tease the fungal sample of the alcohol using a needle mounter, to ensure the sample mixes well with the alcohol.
- 4. Using a dropper or pipette, add one or two drops of Lactophenol Cotton Blue Solution (prepared above) before the ethanol dries off.
- 5. Carefully cover the stain with a clean sterile coverslip without making air bubbles to the stain.
- 6. Examine the stain microscopically at 40X, to observe for fungal spores and other fungal structures.

## **Results and Interpretation**

Fungal spores, hyphae, and fruiting structures stain blue while the background stains pale blue.

#### For example,

- <u>Aspergillus niger</u> stains the hyphae and fruiting structures a delicate blue with a pale blue background.
- <u>Trichophyton</u> mentagrophytes also stains the hyphae and fruiting structures a delicate blue with a pale blue background.

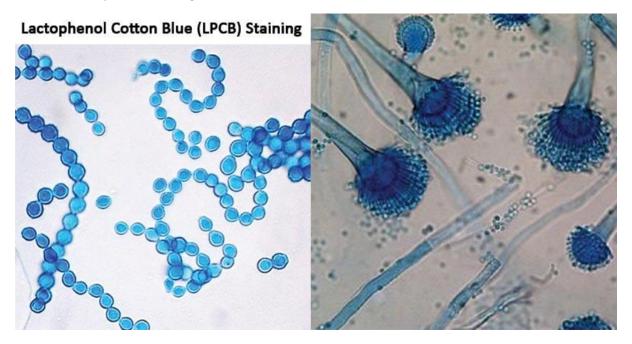


Figure: First: Scopulariopsis species on a lactophenol cotton blue stain. Image

Source: stylish streaking and Senthil Prabhu.

#### Limitations

• It can only be used as a presumptive identification method of fungi which should be followed up with other diagnostic tools such as biochemical and cultural examination.

- The components of the solution should be used before expiry, including the use of the solution before it expires.
- The solution may disrupt the original morphology of the fungi.
- The stain can only be used to identify mature fungi and its structures and not the young vegetative forms of fungi.
- The stain can not be stored for a long period of time.

#### **Applications**

- Used in the identification of suspected fungal samples.
- General identification of fungi and its structures.

#### **References and Sources**

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